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2 January 1997 (02.01.97)

US

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- (75) Inventors/Applicants (for US only): PONCE DE LEON, F., Abel [US/US]; 134 Wildflower Drive, Amherst, MA 10002 (US). CIUFO, Stacy [US/US]; 56 Chesterfield Road, Amherst, MA 01002 (US). ROBL, James [US/US]; 196 Old Enfield, Belchertown, MA 01007 (US). AMBADY, Sakthikumar [IN/IN]; Kerala State (IN). SMYTH, J., Robert, Jr. [US/US]; Amherst, MA 01002 (US).
- (74) Agent: TESKIN, Robin, L.; Burns, Doane, Swecker & Mathis, L.L.P., P.O. Box 1404, Alexandria, VA 22313-1404 (US).

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Published

With international search report.

Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

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(54) Title: Z-CHROMOSOMAL MARKERS DERIVED FROM CHICKEN (GALLUS DOMESTICUS) AND USE THEREOF IN CHROMOSOMAL MAPPING

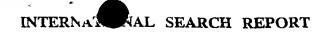
(57) Abstract

We have developed a chicken (Gallus domesticus) Z-chromosome-specific DNA library in a phage vector, by means of chromosome microisolation and microcloning. The chromosomal origin, specificity and purity was evaluated by fluorescent in situ hybridization (FISH) on chicken metaphases. Heterologous chromosome painting, using this Z-chromosome-specific probe on turkey (Meleagris gallopavo) metaphases identified its homologous Z-chromosome, under the same stringent conditions as that used in the chicken, indicating a high degree of Z-chromosome sequence homology among these two species. This chicken Z-chromosome library will facilitate the development of Z-chromosome-specific DNA markers that will be useful for genetic mapping in the domestic chicken and related avian species. The Z-chromosome-specific DNA probe will also be useful for studies pertaining to the sex chromosome evolution in avian species.

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PCT/US 98/08896

A. CLAS	SIFICAT	ION OF	SUBJE	CT MATTER
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According to International Patent Classification (IPC) or to ooth national classification and IPC

8. FIELDS SEARCHED

Minimum gocumentation searched (classification system followed by classification symbols) IPC 6 C120

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
A	LEVIN I ET AL: "Genetic map of the chicken Z chromosome using random amplified polymorphic DNA (RAPD) markers." GENOMICS. (1993 APR) 16 (1) 224-30,	1-7
	XP002067078 cited in the application see the whole document	
A	WO 94 07907 A (ZOOGEN INC) 14 April 1994 see the whole document	1-7
A	WO 96 39505 A (ISIS INNOVATION ;GRIFFITHS RICHARD (GB); TIWARI BELA (GB)) 12 December 1996 see the whole document	1-7

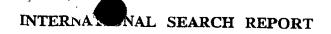
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X Further documents are listed in the continuation of box C.	Patent family members are listed in annex.
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Date of the actual completion of theinternational search 4 June 1998	Date of mailing of the international search report $18/06/1998$
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx, 31 651 epo ni. Fax: (+31-70) 340-3016	Authorized officer Molina Galan, E

INTERNATIONAL SEARCH REPORT



Int. itional Application No PCT/US 98/08896

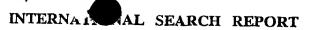
C.(Continue	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category ·	Cilation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Α	BRUFORD M W ET AL: "Minisatellite DNA markers in the chicken genome. II. Isolation and characterization of minisatellite loci." ANIMAL GENETICS, (1994 DEC) 25 (6) 391-9, XP002067079	
A	BIOLOGICAL ABSTRACTS, vol. 95, Philadelphia, PA, US; abstract no. 423269, PONCE DE LEON F A ET AL: "Analysis of the chicken NM7659 T(Z;1) translocation with chromosome painting probes and GBP banding." XP002067083 & EIGHTY-FOURTH ANNUAL MEETING OF THE	
	POULTRY SCIENCE ASSOCIATION, INC., EDMONTON. ALBERTA, CANADA. AUGUST 14-18, 1995. POULTRY SCIENCE 74 (SUPPL. 1). 1995. 9. ISSN: 0032-5791,	
A	BIOLOGICAL ABSTRACTS, vol. 95, Philadelphia, PA, US; abstract no. 423268, AMBADY S ET AL: "A Z - chromosome specific DNA library." XP002067084 & EIGHTY-FOURTH ANNUAL MEETING OF THE	
	POULTRY SCIENCE ASSOCIATION, INC., EDMONTON, ALBERTA, CANADA, AUGUST 14-18, 1995. POULTRY SCIENCE 74 (SUPPL. 1). 1995. 8. ISSN: 0032-5791,	
A	PONCE DE LEÓN ET AL.: "Development of a bovine X chromosome linkage group and painting probes to asses cattle, sheep and goat X chromosome segment homologies" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 93, April 1996, WASHINGTON US, pages 3450-3454, XPO02067080 cited in the application	
P,X	AMBADY S ET AL: "Development of a chicken Z - chromosome -specific DNA library." JOURNAL OF HEREDITY, (1997 MAY-JUN) 88 (3) 247-9, XP002067081 see the whole document	1-7
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	ILION) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication where appropriate, of the relevant passages		Relevant to claim No.
P , X	ZIMMER R ET AL: "Generation of chicken Z - chromosome painting probes by microdissection for screening large-insert genomic libraries." CYTOGENETICS AND CELL GENETICS, (1997) 78 (2) 124-30. XP002067082 see the whole document		1-7
ο, χ	BIOLOGICAL ABSTRACTS, vol. 97, Philadelphia. PA, US: abstract no. 487182,		1-7
•	PONCE DE LEON F A ET AL: "Chicken genome project: Chromosome-specific libraries and applications of genome scans to assess genomic variation." XP002067085 see abstract & REVISTA BRASILEIRA DE REPRODUCAO ANIMAL 21 (3). 1997. 102-105. ISSN: 0102-0803.	· · · · · · · · · · · · · · · · · · ·	
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int. tional Application No PCT/US 98/08896

Information on patent family members

Patent document cited in search repor	t	Publication date		atent family nember(s)	Publication date
WO 9407907	A	14-04-1994	CA AU AU EP	2124220 A 662564 B 2696092 A 0623139 A	14-04-1994 07-09-1995 26-04-1994 09-11-1994
WO 9639505	A	12-12-1996	AU EP	59 0 6996 A 0832218 A	24-12-1996 01-04-1998

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NOTIFICATION OF ELECTION

(PCT Rule 61.2)

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Date of mailing: 27 August 1998 (27.08.98)	in its capacity as elected Office
International application No.: PCT/US98/08896	Applicant's or agent's file reference: 002076-001
International filing date: 02 January 1998 (02.01.98)	Priority date: 02 January 1997 (02.01.97)
Applicant: PONCE DE LEON, F., Abel et al	

1.	The designated Office	e is hereby notified of its	election made:				*	
	X in the demand	filed with the Internation	nal preliminary Ex	amining Autho	rity on:			,
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INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

plicant's or agent's file reference	Sec.	e Notification of Transmittal of International eliminary Examination Report (Form PCT/IPEA/416)
002076-001	FOR FURTHER ACTION Pr	
ernational application No.	International filing date (day/month	hiyear) Priority date (dayimonthiyear)
	02/01/1998	02/01/1997
PCT/US 98/ 08896 ternational Patent Classification (IPC)	or national classification and IPC	
ternational Patent Classification (110)	C12Q1/68	
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UNIVERSITY OF MASSACHU	SEIIS et al.	
Authority and is transmitted to)	y this International Preliminary Examining
2. This REPORT consists of a	total of sheets, including thi	2 COAGL PHEET.
This report is also accombeen amended and are the	panied by ANNEXES, i.e., sheets of t e basis for this report and/or sheets con on 607 of the Administrative Instruction	he description, claims and or drawings which have taining rectifications made before this Authority as under the PCT).
These annexes consists of a to	tal ofsheets.	
3. This report contains indication	is relating to the following items:	
I X Basis of the report		
II Priority	of opinion with regard to novelty, inves	ntive step and industrial applicability
IV Lack of unity of in	venuon	velty, inventive step or industrial applicability;
V Reasoned statement citations and explain	nations supporting such statement	
VI Certain document	s cited	
	the international application	
	ons on the international application	
VIII Certain observation	ons of the international appara	
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	· .	
	Date o	f completion of this report
Date of submission of the demand	Jac v	•
30/07/1998		1 5. 10. 98
	Author	rized officer
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European Patent Office, NL-2280 HV Rijswijk - Tel.: (+31-70) 340-2040		71-20 340 3560
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	١.	Basis	of the	report
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This report has been drawn up on the basis of (Replacement sheets which invitation under Article 14 are referred to in this report as "originally filed" and	have been furnished to the receiving Office in response to an d are not annexed to the report since they do not contain
amendments.)	
the international application as originally filed	
☐ the description, pages	as originally filed
pages	filed with the demand
pages	, filed with the letter of
the claims. Nos.	, as originally filed
Nos.	as amended under Article 19
Nos.	filed with the demand
Nos.	, med was
☐ the drawings, sheets / fig.	as originally filed
sheets / fig.	, filed with the demand
sheets / fig.	, filled with the lower of
2. The amendments have resulted in the cancellation of	
the description, pages:	
the claims. Nos.	
the drawings, sheets / fig.	
3. This report has been established as if (some of) the amendme	nts had not been made, since they have been considered to go
beyond the disclosure as filed (Rule 70.2 (c)).	
4. Additional observations, if necessary:	

national application No.



INTERNATIONAL PRELIMINARY EXAMINATION REPORT

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

	Chahamant
1	Statement

Novelty	Claims	1-7	YES
Novely	Claims		NO
Inventive Step	Claims	1- 7	YES
MAGINIA OCED	Claims		NO
Industrial Applicability	Claims	1-7	YES
Higgs of the same of	Claims		NO

^{2.} Citations and Explanations

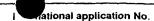
2.1 CITATIONS

Reference is made to the following documents:

D1: Genomics, 16, 1993, 224- 230, Levin et al.

D2: Proc. Natl. Acad. Sci., 93, 1996, 3450- 3454, Ponce de León et al.

- 2.2 NOVELTY (Art. 33(2) PCT)
- 2.2.1 The present application does satisfy the criterion set forth in Article 33(2) PCT because the subject- matter of Claims 1-7 is new in respect of prior art as defined in the regulations (Rule 64(1)- (3) PCT).
- 2.3 INVENTIVE STEP (Art. 33(3) PCT)
- 2.3.1 Document D1, which is considered to represent the most relevant state of the art, discloses (cf. discussion) DNA markers derived from the chicken Z chromosome and methods for using them. The subject- matter of Claim 1 differs in that different markers are claimed.
- 2.3.2 The problem to be solved by the present invention may therefore be regarded as the



PCT/US98/08896

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

provision of alternative DNA markers derived from the chicken Z chromosome. The solution would be the markers identified by Seq. lds. 1-19.

- 2.3.3 Although D2 discloses a method to derive markers from chromosomes similar to that used in the application, it does not seem obvious to derive exactly the markers claimed by the applicant, specially taking into account that the source is a complete chromosome which has not been completely sequenced.
- 2.3.4 For these reasons the markers claimed can not be regarded as a simple choice and the IPEA is of the opinion that the present application satisfies the criterion set forth in Article 33(3) PCT and the subject- matter of claims 1-7 involves an inventive step (Rule 65(1)(2) PCT).

RECORD CUPY

REQUEST

The undersigned requests that the present international application be processed according to the Patent Cooperation Treaty.

International

International Filing Date

Name of receiving Diffice and "PCT Intercalidate Application"

Applicant's or agent's file reference 002076-001 (if desired) (12 characters maximum) Box No. I TITLE OF INVENTION Z-CHROMOSOMAL MARKERS DERIVED FROM CHICKEN (GALLUS DOMESTICUS) AND USE THEREOF IN CHROMOSOMAL MAPPING Box No. II APPLICANT Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (i.e. country) of residence if no State of residence is indicated below.) This person is also inventor. Telephone No. (413) 545-2312 UNIVERSITY OF MASSACHUSETTS A Public Institution of Higher Education of the Commonwealth Facsimile No. of Massachusetts, as Represented by its Amherst Campus (413) 545-6326 Office of Vice Chancellor for Research at Amherst Amherst, Massachusetts 01002 Teleprinter No. United States of America State (i.e. country) of nationality: US State (i.e. country) of residence: US the States indicated in all designated States except the United States This person is applicant all designated the United States of America of America only the Supplemental Box for the purposes of States FURTHER APPLICANT(S) AND/OR (FURTHER) INVENTOR(S) BOX No. III Name and address: (Family name followed by given name; for a legal entity, full official designation. The address must include postal code and name of country. The country of the address indicated in this Box is the applicant's State (i.e. country) of residence if no State of residence is indicated below) This person is: applicant only PONCE DE LEON, F. Abel applicant and inventor 134 Wildflower Drive Amherst, Massachusetts 10002 inventor only (If this check-box United States of America is marked, do not fill in below.) State (i.e. country) of residence: US State (i.e. country) of nationality: US | X | the United States the States indicated in This person is applicant all designated all designated States except the Supplemental Box the United States of America of America only States for the purposes of Further applicants and/or (further) inventors are indicated on a continuation sheet. \mathbf{X} AGENT OR COMMON REPRESENTATIVE; OR ADDRESS FOR CORRESPONDENCE BOX No. IV The person identified below is hereby/has been appointed to act on behalf X agent common representative of the applicant(s) before the competent International Authorities as: Name and address: (Family name followed by given name; for a legal entity, full official Telephone No. (703) 836-6620 designation. The address must include postal code and name of country.) TESKIN, Robin L. Facsimile No. (703) 836-2021 BURNS, DOANE, SWECKER & MATHIS, L.L.P. P.O. Box 1404 Alexandria, Virginia 22313-1404 Teleprinter No. United States of America Mark this check-box where no agent or common representative is/has been appointed and the space above is used instead to .

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Sheet No. 2

T/US 97 / 23 821

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	BY	Belarus	X		Norway
		Canada	IX	NZ	New Zealand
		and LI Switzerland and Liechtenstein	X	PL	Poland
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X	CZ	Czech Republic	LXI	RU	Russian Federation
X	DE	Germany	X	SD	Sudan
X		Denmark	X	SE	Sweden
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	KE	Kenya			
X	KG	Kyrgyzstan	X	UZ	Uzbekistan
X	KP	Democratic People's Republic of Korea	X	VN	Viet Nam
Į			X	YU	Yugoslavia
X	KR	Republic of Korea	X	ZW	Zimbabwe
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X	LT	Lithuania			
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In addition to the designations made above, the applicant also makes under Rule 4.9(b) all designations which would be permitted under the PCT except the designation(s) of The applicant declares that those additional designations are subject to confirmation and that any designation which is not confirmed before the expiration of 15 months from the priority date is to be regarded as withdrawn by the applicant at the expiration of that time limit. (Confirmation of a designation consists of the filing of a notice specifying that designation and the payment of the designation and confirmation fees. Confirmation must reach the receiving Office within the 15-month time limit.)					
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Box No. VI PRIORITY CLA	IM ·	Further priority claims are	e indicated in the Supplemental Box		
The priority of the following earlier application(s) is hereby claimed:					
Country (in which, or for which, the application was filed)	Filing Date (day/month/year)	Application No.	Office of filing (only for regional or international application)		
item (1) US	02 January 1997 (02.01.97)	60/034,410			
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Next to each signature, indicate the name of the person signing and the capacity in which the person signs (if such capacity is not obvious from reading the request). Robin L. Teskin					
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Z-CHROMOSOMAL MARKERS DERIVED FROM CHICKEN (GALLUS DOMESTICUS) AND USE THEREOF IN CHROMOSOMAL MAPPING

Field of the Invention

The invention relates to novel chromosomal markers derived from chicken and use thereof.

Background of the Invention

Livestock genome maps have progressed very rapidly in the past few years due to the availability of highly polymorphic DNA markers. But in many species, the maps are not dense enough to facilitate a thorough search for quantitative trait loci (QTLs). This is especially true in the case of the chicken. The chicken haploid karyotype consists of 39 chromosomes that are classified into two categories - the macrochromosomes and the microchromosomes. The largest five pairs of macrochromosomes and the Z-chromosome represent about 55 percent of the total DNA content of the chicken genome. The Z-chromosome covers about 210 cM of the estimated 2500 - 3,000 cM of the chicken genome map (Levin et al. *Genomics*, 16:224-230 (1993)).

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Knowledge of the genetic composition of the chicken Z-chromosome is limited, in spite of the fact that this chromosome has the most detailed linkage map for this species, largely generated by classical linkage test analyses (Bitgood and Somes, *Poultry Breeding and Genetics*, 2nd Ed., Crawford RD, ed., Amsterdam: Elsevier, pp. 469-495 (1990)). To date, 19 known loci and 14 genetic markers consisting of 3 chicken middle repetitive sequence element (CRI) markers, 8 random amplified polymorphic DNA (RAPD) markers and 3 microsatellites have been assigned to the chicken Z-chromosome (Bitgood and Somes, (Id.) (1990); Saitoh et al, *Chrom. Res.*, 1: 239-251 (1993); Cheng et al, *Poultry Sci.*, 74: 1855-1874 (1995)).

The avian sex chromosome constitution differs from that of mammals because females are heterogametic (ZW) and males homogametic (ZZ). It has been observed from comparative linkage analyses that some of the sex linked

genes in mammals are autosomal in chicken, while some of the sex linked genes in chicken are autosomal in mammals (Bitgood and Somes, (Id.) (1990)). Accordingly, obtaining further information concerning the Z-chromosome of chickens would be beneficial in identifying sex-linked genes in chickens and related species.

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Brief Description and Objects of the Invention

Thus, it is an object of the invention to identify novel chromosomal markers from the Z-chromosome of chicken. It is further an object of the invention to use such markers to construct a Z-chromosome specific DNA map and to use such chromosomal markers to identify Z-chromosome homologs in related avian species, e.g., turkey.

In order to develop a dense genetic map for chicken, it is important to generate a large number of polymorphic markers per chromosome (Cheng et al, *Poultry Sci.*, 741:1855-1874 (1995)). One way of achieving this goal is to develop chromosome-specific libraries. Chromosome flow-sorting has been the method of choice for the generation of chromosome-specific libraries in humans (Fuscoe et al, *Cytogenet Cell Genet*, 43:79-86 (1986)) and in swine (Langford et al, *Anim. Genet*, 24: 261-267 (1993)). Development of flow-sorted chromosomes is technically demanding and frequently yield preparations which have some degree of contamination with other chromosomes (Hozier and Davis, *Anal. Biochem*, 200: 205-127 (1992)).

A more effective and direct way of generating chromosome-specific DNA libraries is by chromosome microisolation and microcloning of the chromosome of interest. Chromosome specific libraries generated by chromosome microisolation have been used in swine (Ambady et al, (unpublished data)), cattle (Ponce de León et al, *Proc. Natl. Acad. Sci., USA*, (in press) 1996)), and chicken (Li et al, *Proc. of the 10th Eur. Colloq. on Cytogenetics of Domestic Animals*, Utrecht Univ., The Neth., p. 11, August 18-21 (1992)) genetic mapping studies in order to develop maps for particular chromosomes.

Generation of polymorphic markers from chromosome-specific libraries for all of the 8 pairs of the chicken macrochromosomes will enable saturation of about 55-70% of the chicken genome. Chromosome-specific DNA can also be used as heterologous chromosome painting probes in closely and distantly related species for comparative genome analysis, study of chromosomal evolution, and for identifying gross chromosomal abnormalities.

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This application, in particular, provides a chicken Z-chromosome-specific DNA library, Z-chromosomal markers and use thereof as probes to identify the Z-chromosome homolog in related species, such as turkey.

Brief Description of the Figures

Figure 1 shows amplification of microsatellite markers by PCR and identification of polymorphisms.

Figure 2 shows a genetic map constructed using the identified microsatellite markers.

Figure 3 shows dinucleotide repeats present in the identified microsatellite markers.

Detailed Description of the Invention

Microisolation and microcloning:

Chicken metaphases were prepared from chicken fibroblast cultures

following standard procedures, fixed briefly for 5 minutes each in 9:1, 5:1 and
3:1 methanol:acetic acid and dropped on clean coverslips. Chromosome
microisolation and cloning was performed following the procedure described by
Ponce de León et al (*Proc. Natl. Acad. Sci., USA* (in press) (1996)). Briefly,
twelve copies of the chicken Z-chromosome were microisolated and transferred
to clean siliconized coverslips. Proteinase-K digestion, phenol-chloroform
extraction, *Sau*3AI (50U/μl, New England Biolabs) digestion and ligation to
custom prepared *Sau*3AI adaptors were performed in a nanoliter drop. Ligation

products were digested with BgII enzyme (Promega, 10 units/ μ l) to cleave off the adaptor dimers that form during the ligation process.

The ligation product was PCR amplified and 10 μ l of the amplified product was run on an agarose gel to determine the size of the amplified products. A 2 μ l volume of this original amplification was labeled by PCR, using biotin-16-dUTP (Boehringer Mannheim). The purity, specificity and origin of the DNA fragments was determined by FISH on chicken metaphases following the procedure described by Ponce de León et al (*Proc. Natl. Acad. Sci. USA* (in press) (1996)). The remainder of the PCR product was digested with *Sau*3AI and passed through a Microcon 30 (Amicon Inc.) spin column to cleave and remove the flanking adaptors respectively.

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In order to produce a chicken Z-chromosome-specific phage library, the digested DNA was cloned in a lambda ZAP Express vector (Stratagene) and packaged using Gigapack II Gold packaging extract (Stratagene). The library was amplified by plate lysate method following the manufacturer's protocol and stored at -70°C in 7% DMSO and 0.3% chloroform. Average size of library inserts was determined by PCR amplification of 30 randomly picked clones using the T3 and T7 priming sites flanking the insert.

Fluorescent in situ hybridizations

20 The Z-chromosome-specific DNA fragments were fluorescently labeled by PCR with biotin-16-dUTP (3:1 ratio of dTTP:biotin-16-dUTP) and passed through a Sephadex G-50 column to remove unincorporated nucleotides. The protocol described by Ponce de León (*Proc. Natl. Acad. Sci., USA* (in press) (1996)) was followed. Briefly, 200 nanograms of labeled Z-chromosome specific DNA was mixed with 6 μg of chicken competitor DNA (average size 200-400 bp) and 5.8 μg of salmon sperm DNA (average size 200-400 bp), precipitated and resuspended in 12 μl of hybridization buffer consisting of 50% deionized formamide, 1X SSC and 100% dextran sulphate to achieve a final DNA concentration of 1 μg/μl. The hybridization mix was denatured at 75°C for 5

minutes and reannealed at 37°C for 10 minutes and deposited on denatured (70% formamide, 2X SSC at 70°C for 2 minutes) chicken or turkey metaphases, mounted, sealed with rubber cement and incubated in a humidified chamber at 37°C for 18 to 20 hours. The slides were washed in 50% formamide/2X SSC at 42°C for 15 minutes and 0.1X SSC at 60°C for 15 minutes. Blocking was done using 2% blocking reagent (Boehringer Mannheim) and the signals were detected using avidin-FITC (5 μ g/ml, Vector labs) in 1% blocking solution. Slides were washed in 4X SSC/0.1% Tween-20 for 15 minutes at 42°C, stained for 10 minutes in propidium iodide (400 ng/ml in 2X SSC) and rinsed for 5 minutes in 2X SSC/0.01% Tween-20. Slides were mounted in p-phenylenediamine-11 (PPD-11) antifade and observed under a Zeiss Axioskop fluorescent microscope.

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Results

A chicken Z-chromosome specific DNA cocktail was developed by chromosome microisolation, Sau3AI digestion, adaptor ligation and PCR amplification. The amplified DNA fragments ranged in size from 400 bp to 1600 bp with the bulk of the DNA in the 500-1000 bp range. The origin, specificity and purity of the chromosomal DNA fragments was verified by FISH after PCR labeling of a small fraction of the DNA cocktail. The probes showed specific hybridization signal on a medium sized submetacentric chromosome identified as the Z-chromosome based on its morphology and G-banding pattern. After having confirmed the origin and purity of the preparation, the adaptors flanking the inserts were removed by Sau3AI digestion and column purification. Cloning was performed using equimolar ratios of the inserts to the vector ends (lambda ZAP Express, Stratagene). The original library consisted of a total of 8.48 X 10⁵ plaques representing about 14 chicken Z-chromosome equivalents. The final titer of the amplified library was 1.2 X 10¹² pfu/ml.

Thirty random plaques were selected and the inserts PCR-amplified using the T3/T7 priming sites flanking the inserts. The average insert size was about 1,000 bp (data not shown). This library was screened to identify microsatellite

containing clones to increase the marker density of the chicken Z-chromosome genetic linkage map.

Heterologous painting of turkey metaphase chromosomes:

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The labeled chicken Z-chromosome-specific DNA fragments were used to perform FISH analysis on turkey metaphase chromosomes following the procedure described previously. Washes at the same stringency showed strong hybridization signals on a medium-sized submetacentric chromosome in turkey metaphases (data not shown). This chromosome was identified as the Zchromosome homolog in the turkey. The obtained results indicate that the chicken and turkey Z-chromosome sequences are highly conserved. The redlegged partridge Z-chromosome has also been shown to be homologous to the chicken Z-chromosome (Dias el al, Proc. of the XXIV Int. Cont. on Anim. Genet., Prague, Czech. p. 133 (July 23-24, 1994)). These results are similar to the FISH results obtained when the bovine X-chromosome painting probes were used on sheep and goat chromosomes (Ponce de León el al, Proc. Natl. Acad. Sci., USA (in press) (1996)) and with human X-chromosome probes on a wide range of mammalian species (Schertan et al, Nat. Genet., 6:342-347 (1994)) indicating the high degree of sex chromosome conservation among all the mammalian species studied. Solinas-Toldo et al (Genomics, 27: 489-496 (1995)) have previously shown that human chromosome-specific painting probes could identify chromosomal segments in bovine that are homologous to specific human chromosomes. It is expected based on our results that chicken chromosome painting probes can similarly be used in closely and distantly related avian species to identify gross chromosomal rearrangements such as translocations and duplications that have occurred during avian evolution. Since the chicken Zchromosome sequences are highly conserved in the turkey, the chicken Zchromosome-specific microsatellite markers should be particularly useful for genetic mapping in turkey.

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Conclusions

Genetic and physical mapping of human and animal genomes has been greatly facilitated by the use of chromosome specific DNA libraries. Mapping with libraries specific to a chromosome or chromosomal region increases marker saturation by reducing the gaps resulting from a purely random shotgun 5 approach. This study was undertaken to construct a genetic and physical map of microsatellites on the chicken Z chromosome. This chromosome is the fifth largest in the chicken genome, comprising about 8% of the total. Notwithstanding its size, very few microsatellites have been assigned to it. DNA originating from the chicken Z chromosome was previously isolated and 10 reported. This was used to construct a small insert library in Lambda ZAP Express, representing 14 chromosome equivalents. This library was screened for microsatellites with an (AC) 12 oligo, and positive clones were isolated. Confirmation of the presence of the microsatellite, as well as its approximate location along the cloned fragment was accomplished by PCR amplification. 15 Clones with adequate flanking regions were sequenced, and primers for 19 microsatellites were constructed. These primers were used to genotype individuals from the East Lansing Poultry Reference Population and a linkage map was constructed. Fourteen markers were scorable and polymorphic in this population. The resulting map contains 12 markers in two linkage groups span-20 ning 90 Cm and two unlinked markers. The physical location of each marker was established by fluorescent in situ hybridization (FISH). Preliminary results with four markers allowed the assignment of one linkage group to the long arm of the Z chromosome, and one to the short arm.

The following nucleic acid sequences are microsatellite markers identified by the above methods. As discussed supra, these markers are useful for genetic mapping and for study of the sex chromosome structure in avian species. Also, such markers should enable the identification of genes encoding desirable traits,

e.g., genes involved in growth rates, and for identifying sex-linked genotypes.

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EXAMPLE

The specific <u>Gallus domesticus</u> microsatellite markers identified are set forth below. As noted, these DNA markers will be useful for genetic mapping of domestic chicken as well as related avian species and for studies pertaining to evolution of the sex chromosome in avian species.

SEQUENCE 1 (43. Seq)

- 1 gatcactttc cctaatattc ttgtgtttct tgtttgttga cctgtaatgc
- 1 agttctgagt tttggaaagg aactaattaa gaccagagga gagataattt
- 101 tettttatea aaaaacaaac aaacaaacaa aaaaacgaat tettaceact
- 10 151 ttacaaaaat tttccatttt gaaggccagt acagccatag cattcatcta
 - 201 ctttttgctt tggat

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<u>SEQUENCE 2</u> (71. Seq)

- 1 gatcaggtgg cctgtagtag acaacaacaa caatggggtg ccctttgttg
- 51 cettagtete taactegeae ceaeacaea ttteaagttg ettgtggeea
- 15 101 ttetteaggg acagttette acaatetatt cettteetga tgtagaagge
 - 151 gteacetect ecceteetge etegtttgte eettetaaac tgeaggtatt
 - 201 agtattgata getaaggtea agteatggga accateteae eaggttteag
 - 251 tgttggcaac tatgttatgc tttcttagga gcatggtggt tccaactctt
 - 301 ccctgcttat ttcccaagct gtgtgtgatg gtaggatagc attcaagtgg
- 20 351 gaggagceta teggettttt ggaggtaete etaaateeet gatatteeee
 - 401 tgattcccgt acttcttcct tgccaagggc ccgccaatgc atagttcaat
 - 451 ttctcatgca gacgctaagg aaaggtggac cc

SEQUENCE 3 (80 Seq.)

- 1 gategtatgt attttttac ataggataga aaatggccaa taggaaataa
- 51 gacagtacag ctactaagaa agaaacacaa ttacacacac acacacacac
- 101 acacacaca acacatttga aaaacgcgct gcacagcagt gtgggtattt
- 5 151 tttcacaaga gagacacact ctacagtaca cagccagctc tactttgtcg
 - 201 cacagtetea gtgtgtgttt gecaacagga egeggtteae agggagatat
 - 251 tgtcctcttg tgtgtgtgga gacacagaga cagag

SEQUENCE 4 (81. Seq)

- 1 gateceetgg aggaagggea atggeaacce acteeagtat tettgeetga
- 10 51 agaataccat ggtcagtttt gcctcctggg ctatagtcca tggggttgca
 - 101 aagagteagg eatgactgag egactetete tetetetete tetetetete
 - 151 acacacaca acacacaca acacacggcg tetetetete tetetataca
 - 201 tataggetgt gtgteteget atteteaeat gagggaaaet catatetage
 - 251 acgtggcaca aatattgttt gtggctctca caaaagacat gtgggcgcac
- 15 301 aaaggteeec eeceggtgga taeanegeet tggtttttta taaceeaage
 - 351 ctgtg

SEQUENCE 5 (131 Seq)

- 1 gatcacatat gtaaactagg gaattgcata ataagattaa atgtaggtgt
- 51 agaacgtggc atgaaggaag gtagaattag gtggtaccta tetettetga
- 20 101 aacaaactga gaatectact accaatcaac atattetaca taccacacac
 - 151 acattttttc tcgagtaaaa tataaactaa tgagaaactt ccctag

SEQUENCE 6 (147. Seq)

1 gateceaage aacacatagn cagacaatea cacacacaca cacacacaca
51 cacacacaca cacacacaca cacateetet eeceacaata cateeegaga
101 ggggggagag acactetete teeeteta taggggagae eeggagaget
5 151 ggetetgttg tetetetaca eeggacatac agtggageac ateteacact
201 tgtgtetttg tetetetaca eeggacatac agtggageac ateteacact
251 tgtgteteta teteteeetg teeetgttga teeatetete tteacacate
301 tetecagate ttagegetag agteteetgt ettetetetg egcaatttgt
351 gtgatagaga cacetgatat gttgtgtggg ggagacatet gtgtgtetet
10 401 gtgteateee agaggatttt teteteecac aettagagge etteteaaga
451 gatgggaggt tttaatgggg tgtg

SEQUENCE 7 (166. Seq)

- 1 gatcattett etgttteeea ttetaatggg aatteteeae acacacaca
- 51 acacacaca acacacacat ettetteece ttacatggaa aaaaateete
- 15 101 cacaccctg gacactgatt actetecete tteecagaga gagate

SEQUENCE 8 (196. Seq)

- 1 gateceetag agaagggaat ggetaeteae teeagtatte ttgeetggag
- 51 aatteegtgg teagaggage etggaagget ataateeata gagtegeaag
- 101 agtcagacag gactgagtga ctaacacaca catgcacaca cacacacaca
- 20 151 cacacacaca cttgctctag ggagaggcat agagatgtaa tctctcctaa
 - 201 aatggggtg gegatggeee etgeggeeaa gtaategeea caeatgegta
 - 251 tteeeettaa gattgggtta ggeeteeett atgaggagag aecagggaga
 - 301 gaatgggete tetetetete teaeteecea accgagtaag tggtaaaaaa
 - 351 ggttttcctg gattacaatt ttggtgttac agaattggaa aaaaatattt
- 25 401 ttggggetee eeeteagtt ta

SEQUENCE 9 (199. Seq)

- 1 ctagcaaaaa caccccaca agttatgaaa acaacggctt aatatagtaa
- 51 tgtgtgtgtg tgtgtgtgtg tgttgcacac cacagttttc tctgatactc
- 101 aaacetetet etttetetae aggggeeece cataacacag eggetgagat
- 5 151 gtgtgacggg aaggcgtggc cttttacaca tttgtggtat ggtctgccaa
 - 201 ggcccctat tgcccccac aactacggag atacactagg ggcgacccgc
 - 251 aggegegega ecceeaggtg gggeeegag

SEQUENCE 10 (204. Seq)

- 1 ctttaggagg ttctctcgag taagcttttt ggatttcttt ggttcccaag
- 10 51 catcacatgg tacaggcagt cacacacaca cacatacaca cacacacaca
 - 101 cacacacaca cactectete eccacaatae atacegagag gggggagaga
 - 151 cactetetet eeetetetat agggggagee eeacagaget ggetetgttg
 - 201 teteteteea eeggacatae agtggageae ateteaeaet tetgteteta
 - 251 tetetecetg eccetgtgae atecatetet etteacacaa teteacecag
- 15 301 gatettageg etagagaeee eetgteette tteteetggg gaaatttttt
 - 351 gtggataaga gacacccgat atattggtgt gggggagaac atcttgtgag
 - 401 gtetetgttg tgecatecea acaggaattt ttateteece cacaattaga
 - 451 ggcccccct caagagtgtg tgagggtt

SEQUENCE 11 (235. Seq)

- 20 1 gatcacagat gtatgtattt ttttacatag gatagaaaat ggacaatagg
 - 51 aaataagaca gtacagctac taagaaagaa cccacattta cacacacaca
 - 101 cacacacaca cacacacaca agtgtttaat cegetgeaca geattgtgga
 - 151 catttttaca caagagagac acactetaca gtttgcgccc agetetag

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SEQUENCE 12 (249. Seq.)

- 1 gatcattett etgttteeca ttetaatgga atteteeaca eacaeacaea
- 51 cacacacaca cacacactet tettteteet gacatggaaa aateteeece
- 101 acacceggg acactgattt etetecetet ecceaacaet gtgagcaaga
- 5 151 ggagtttatt ttgtgtgtgt cactetteca gggagagaga gate

SEQUENCE 13 (258. Seq)

- 1 ctaggeateg gttgggaggt ggtgagtaat tacttgtetg acattagtee
- 51 tgtaacattg ggtgtgtgt tgtgtgtgtg tgtgtattcc ccttgggaat
- 101 tggttttete aaccacaagt tettettttt tttttttete eeceetttte
- 10 151 ttctgaaaat aagtacttgg ggggtttccg ccccccgg taaataaaat

SEQUENCE 14 (290. Seq)

- 1 ctagtggete ccaagcaaca catagecaga caacacacac acacacaca
- 51 acacacaca acacacaca acacacacte etetececae aatacateee
- 101 gagaggggg agagacacte tetetecete tetatagegg gagececaca
- 15 151 gagetggete tgetgtetet etacaeegga eataeagtgg ageaeatete
 - 201 acattegtgt etetatetet eeetgeeeet ggtgacatae atetetette
 - 251 acacatetea ceaggtetga gegetagagt etcetgtett etetetgege
 - 301 aatatttgtg atagagacat ctgatatatt gtgtgtggga gacatcttgt
 - 351 gagtetetgt gtgeatecea gaggattttt ateteeeae aetag

98 0336

SEQUENCE 15 (309. Seq)

1 gatecatgaa aacttteega gttgtattgt etaggtgaaa acacacacaa
51 acacacacac acacacaca acacaacagg gagatgagte ttgcaagaga
101 ataggggaga gttatgteac caagtetggt gaggtatata gegtataggg
151 agceaacatg teagacatet gatgtgetaa gattaacatt ttattttatt
201 taatgtgtga gateteatat ageggetett ettatatatg aegtetegea
251 atgtetettt atgtgtgtta ttetetgage eeetgggaga tatetgteat
301 cagagagaag agacatacac atacaggggt tatatatttt eteeetgtgt
351 gtggagatgg agggtatttt ggacaagete aacacteatt ggeteecaga

401 gagagaaaag gagcaactgt tgcacccggg gctctgtagc tgggatc

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SEQUENCE 16 (341. Seq)

1 caattgggta catctacctg gtaccccace cgggtggaaa atcgcatggg
51 cccgcggcgg ttctaggaag tactctcgag aagcttttgg gttctttggg
101 tcccaagcag cacatggaca ggcaatcaca cacacacaca cacacacaca
151 cacacacaca cacacacaca ctcctctccc cacaatacat cccgagaggg
201 gggagagtca ctctctctcc ctctctatag ggggcgcccc taagagctgg
251 ctctgttgtc tatctacacc gcacatacaa tggagcacaa ctcacactag

SEQUENCE 17 (398. Seq)

1 gatcaaagca tggaggtcat gccaggcact gaacaaaatg gtagagagtg
20 51 attctatgac tgactaagac ctcatgcaac aacaagtgaa gagtcacaac
101 tgcaaacaga agtacaactt agcaaatcct attttcagga aacactaaac
151 cgtaatactt gcacgatttt ttctttaata cagtaataat tcttttagaa
201 tttggatata tcttttaaga tacatatttg tctaaatacc aaggcaggat
251 atgagcataa aatagctaag gttagctatg gtgttatatt taagaagacc
25 301 acagagcaat aggagcatac ttttcttggg gtagaagggg cccttaaagg
351 tcacctag

SEQUENCE 18 (420. Seq)

- 1 ctagccacat cctataacte cactccacct ttaatcctga tttctgtgtc
- 51 tettetetaa eetetatgge etttetetaa agtteeceaa tateaacaat
- 101 cetttteece aetgggaeet eeagtttatt gattetaeea tgteaetate
- 5 151 catggtcaac cacttgtggt attataggat gtcgcgtgtg tgtgtgtgtg
 - 201 tgtgtgcatg tgtgtgtgct tgggtgtcag agagttccaa tctgggggac
 - 251 ctatggtttg taaacaacag gtctcttgcc aaggaagat

SEQUENCE 19 (435. Seq)

- 1 ctagegeteg tgeceetgea gttegacaet eagtggetee tecacaeaea
- 10 51 cacacacaca cacatcaata tatatataga tagatagata gatagaggag
 - 101 caatataagt ggetteteta ttteeageat gttttgaaga geataaacte
 - 151 aacagagtat atataaatet gatgtgaccc atgtcatetg ctacagcatg
 - 201 agagggggta gtgatc

CLAIMS:

- 1. A Z-chromosomal marker DNA selected from the group consisting of Sequence I (43. Seq), Sequence 2 (71. Seq), Sequence 3 (80. Seq), Sequence 4 (81. Seq), Sequence 5 (131. Seq), Sequence 6 (147. Seq), Sequence 7 (166. Seq), Sequence 8 (196. Seq), Sequence 9 (199. Seq), Sequence 10 (204. Seq), Sequence 11 (235. Seq), Sequence 12 (249. Seq), Sequence 13 (258. Seq), Sequence 14 (290. Seq), Sequence 15 (309. Seq), Sequence 16 (341. Seq), Sequence 17 (398. Seq), Sequence 18 (420. Seq), and Sequence 19 (435. Seq).
- 2. A Z-chromosomal DNA library that contains at least one DNA sequence according to Claim 1.
 - 3. A method of using at least one Z-chromosomal DNA according to Claim 1 for genetic mapping.
 - 4. The method of Claim 3, wherein the genetic mapping is effected to construct a Z-chromosome specific DNA map.
- 15 5. The method of Claim 3, wherein the Z-chromosome DNA map is that of an avian species selected from the group consisting of chicken, turkey, partridge, duck, guinea hen, and goose.
 - 6. The method of Claim 4, which is used to identify gross chromosomal rearrangements.
- 7. The method of Claim 6, wherein said chromosomal rearrangement comprises a translocation, deletion or duplication.

ABSTRACT

We have developed a chicken (Gallus domesticus) Z-chromosome-specific DNA library in a phage vector, by means of chromosome microisolation and microcloning. The chromosomal origin, specificity and purity was evaluated by fluorescent in situ hybridization (FISH) on chicken metaphases. Heterologous chromosome painting, using this Z-chromosome-specific probe on turkey (Meleagris gallopavo) metaphases identified its homologous Z-chromosome, under the same stringent conditions as that used in the chicken, indicating a high degree of Z-chromosome sequence homology among these two species. This chicken Z-chromosome library will facilitate the development of Z-chromosome-specific DNA markers that will be useful for genetic mapping in the domestic chicken and related avian species. The Z-chromosome-specific DNA probe will also be useful for studies pertaining to the sex chromosome evolution in avian species.

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FIGURE

2/4

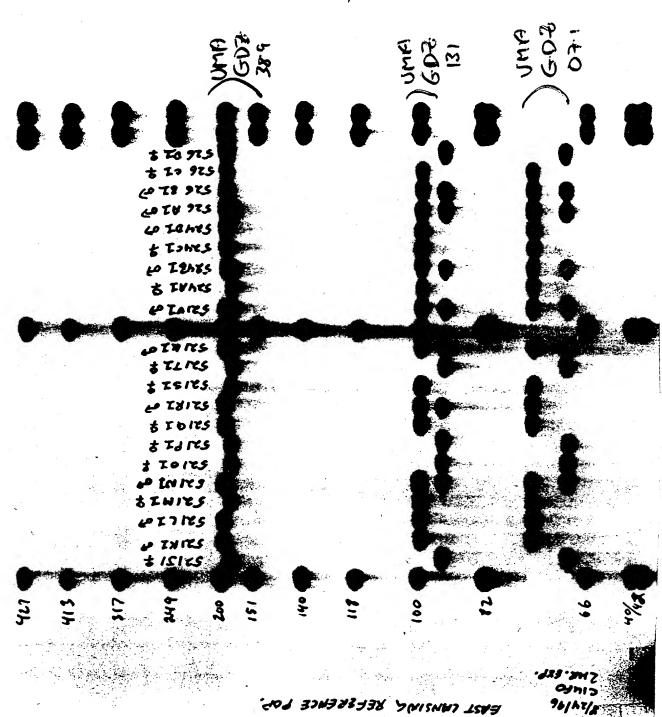
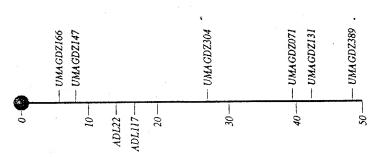


FIGURE 1 (Cont)

FIGURE 2



1:314 GDZ080 PM4GDZ249

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- DMAGDZ081



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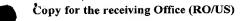
Chicken Z Chromosome Microsatellites Microsatellite composition

s. Ciufo

Clone	Repeat
UMGDZ043	(AAC) ₇
UMGDZ071	(CA) ₅
UMGDZ080	(AC) ₁₆
UMGDZ081	$(CT)_{13}(AC)_{13}(CT)_{7}$
UMGDZ131	(CA) ₄
UMGDZ147	(CA) ₂₂
UMGDZ166	(AC) ₁₅
UMGDZ196	(AC) ₁₉
UMGDZ199	(GT) ₁₂
UMGDZ204	(AC) ₂₁
UMGDZ235	(AC) ₁₅
UMGDZ249	$(AC)_{16}(TTC)_4$
UMGDZ258	(TG) ₁₂
UMGDZ290	(AC) ₂₃
UMGDZ304	(AC) ₂₀
UMGDZ341	(AC) 22
UMGDZ398	(CAA) ₃
UMGDZ420	(GT) ₂₀
UMGDZ435	(CA) ₁₁

FIGURE 3

5 Ho 5 7 6 18 1/95





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International application No.	International filing date (day/month/year)			
PCT/US98/08896	02 January 1998 (02.01.1998)			
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A copy of this notification has been sent to the receiving Office (RO/US) and the International Searching Authority (ISA/EP).				
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The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland	Authorized officer . Addae-Ruesch			

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The applicant is hereby notified that the International Bureau has received the record copy of the international application as detailed below.

Name(s) of the applicant(s) and State(s) for which they are applicants:

UNIVERSITY OF MASSACHUSETTS (for all designated States except US) PONCE DE LEON, F., Abel et al (for US)

International filing date

02 January 1998 (02.01.98)

Priority date(s) claimed

02 January 1997 (02.01.97)

Date of receipt of the record copy by the International Bureau

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This International Search Report has be according to Article 18. A copy is being t	•	chority and is transmitted to the applicant
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A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12Q1/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols) IPC $\,6\,$ C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT				
Category °	Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.	
A	LEVIN I ET AL: "Genetic map of the chicken Z chromosome using random amplified polymorphic DNA (RAPD) markers." GENOMICS, (1993 APR) 16 (1) 224-30, XP002067078 cited in the application see the whole document		1-7	
A	WO 94 07907 A (ZOOGEN INC) 14 April 1994 see the whole document		1-7	
А	WO 96 39505 A (ISIS INNOVATION ;GRIFFITHS RICHARD (GB); TIWARI BELA (GB)) 12 December 1996 see the whole document		1-7	
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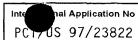
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Ρ,Χ	ZIMMER R ET AL: "Generation of chicken Z – chromosome painting probes by microdissection for screening large-insert genomic libraries." CYTOGENETICS AND CELL GENETICS, (1997) 78 (2) 124-30, XP002067082/see the whole document	1-7	
Ρ,Χ	BIOLOGICAL ABSTRACTS, vol. 97, Philadelphia, PA, US; abstract no. 487182, PONCE DE LEON F A ET AL: "Chicken genome project: Chromosome-specific libraries and applications of genome scans to assess genomic variation." XP002067085 see abstract & REVISTA BRASILEIRA DE REPRODUCAO ANIMAL 21 (3). 1997. 102-105. ISSN: 0102-0803,	1-7	
		-	
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C.(Continua Category °	Citation of document, with indication, where appropriate, of the relevant passages BRUFORD M W ET AL: "Minisatellite DNA markers in the chicken genome. II.	Relevant to claim No.
	BRUFORD M W ET AL: "Minisatellite DNA	 Relevant to claim No.
А		
	Isolation and characterization of minisatellite loci." ANIMAL GENETICS, (1994 DEC) 25 (6) 391-9, XP002067079	
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A	BIOLOGICAL ABSTRACTS, vol. 95, Philadelphia, PA, US; abstract no. 423268, AMBADY S ET AL: "A Z - chromosome specific DNA library." XP002067084 ✓	
	& EIGHTY-FOURTH ANNUAL MEETING OF THE POULTRY SCIENCE ASSOCIATION, INC., EDMONTON, ALBERTA, CANADA, AUGUST 14-18, 1995. POULTRY SCIENCE 74 (SUPPL. 1). 1995. 8. ISSN: 0032-5791,	
A	PONCE DE LEÓN ET AL.: "Development of a bovine X chromosome linkage group and painting probes to asses cattle, sheep and goat X chromosome segment homologies" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, vol. 93, April 1996, WASHINGTON US, pages 3450-3454, XPO02067080 cited in the application	
Ρ,Χ	AMBADY S ET AL: "Development of a chicken Z - chromosome -specific DNA library." JOURNAL OF HEREDITY, (1997 MAY-JUN) 88 (3) 247-9, XP002067081 see the whole document	1-7
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INTERNATIONAL SEARCH REPORT

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Internal Application No PC 17 US 97/23822

Patent document cited in search report				atent family nember(s)	Publication date
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WO 9639505	A	12-12-1996	AU EP	5906996 A 0832218 A	24-12-1996 01-04-1998

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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A1

(11) International Publication Number:

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US

(71) Applicant (for all designated States except US): UNIVERSITY OF MASSACHUSETTS, A PUBLIC INSTITUTION OF HIGHER EDUCATION OF THE COMMONWEALTH OF MASSACHUSSETS, as represented by ITS AMHERST CAMPUS [US/US]; Office of Vice Chancellor for Research at Amherst, Amherst, MA 01002 (US).

(72) Inventors; and

- (75) Inventors/Applicants (for US only): PONCE DE LEON, F., Abel [US/US]; 134 Wildflower Drive, Amherst, MA 10002 (US), CIUFO, Stacy [US/US]; 56 Chesterfield Road, Amherst, MA 01002 (US). ROBL, James [US/US]; 196 Old Enfield, Belchertown, MA 01007 (US). AMBADY, Sakthikumar [IN/IN]; Kerala State (IN). SMYTH, J., Robert, Jr. [US/US]; Amherst, MA 01002 (US).
- (74) Agent: TESKIN, Robin, L.; Burns, Doane, Swecker & Mathis, L.L.P., P.O. Box 1404, Alexandria, VA 22313-1404 (US).

(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published

With international search report.

Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

(54) Title: Z-CHROMOSOMAL MARKERS DERIVED FROM CHICKEN (GALLUS DOMESTICUS) AND USE THEREOF IN CHROMOSOMAL MAPPING

(57) Abstract

We have developed a chicken (Gallus domesticus) Z-chromosome-specific DNA library in a phage vector, by means of chromosome microisolation and microcloning. The chromosomal origin, specificity and purity was evaluated by fluorescent in situ hybridization (FISH) on chicken metaphases. Heterologous chromosome painting, using this Z-chromosome-specific probe on turkey (Meleagris gallopavo) metaphases identified its homologous Z-chromosome, under the same stringent conditions as that used in the chicken, indicating a high degree of Z-chromosome sequence homology among these two species. This chicken Z-chromosome library will facilitate the development of Z-chromosome-specific DNA markers that will be useful for genetic mapping in the domestic chicken and related avian species. The Z-chromosome-specific DNA probe will also be useful for studies pertaining to the sex chromosome evolution in avian species.

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WO 98/37243 PCT/US98/08896

Z-CHROMOSOMAL MARKERS DERIVED FROM CHICKEN (GALLUS DOMESTICUS) AND USE THEREOF IN CHROMOSOMAL MAPPING

Field of the Invention

The invention relates to novel chromosomal markers derived from chicken and use thereof.

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Background of the Invention

Livestock genome maps have progressed very rapidly in the past few years due to the availability of highly polymorphic DNA markers. But in many species, the maps are not dense enough to facilitate a thorough search for quantitative trait loci (QTLs). This is especially true in the case of the chicken. The chicken haploid karyotype consists of 39 chromosomes that are classified into two categories - the macrochromosomes and the microchromosomes. The largest five pairs of macrochromosomes and the Z-chromosome represent about 55 percent of the total DNA content of the chicken genome. The Z-chromosome covers about 210 cM of the estimated 2500 - 3,000 cM of the chicken genome map (Levin et al. *Genomics*, 16:224-230 (1993)).

Knowledge of the genetic composition of the chicken Z-chromosome is limited, in spite of the fact that this chromosome has the most detailed linkage map for this species, largely generated by classical linkage test analyses (Bitgood and Somes, *Poultry Breeding and Genetics*, 2nd Ed., Crawford RD, ed., Amsterdam: Elsevier, pp. 469-495 (1990)). To date, 19 known loci and 14 genetic markers consisting of 3 chicken middle repetitive sequence element (CRI) markers, 8 random amplified polymorphic DNA (RAPD) markers and 3 microsatellites have been assigned to the chicken Z-chromosome (Bitgood and Somes, (Id.) (1990); Saitoh et al, *Chrom. Res.*, 1: 239-251 (1993); Cheng et al, *Poultry Sci.*, 74: 1855-1874 (1995)).

The avian sex chromosome constitution differs from that of mammals because females are heterogametic (ZW) and males homogametic (ZZ). It has been observed from comparative linkage analyses that some of the sex linked

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genes in mammals are autosomal in chicken, while some of the sex linked genes in chicken are autosomal in mammals (Bitgood and Somes, (Id.) (1990)). Accordingly, obtaining further information concerning the Z-chromosome of chickens would be beneficial in identifying sex-linked genes in chickens and related species.

Brief Description and Objects of the Invention

Thus, it is an object of the invention to identify novel chromosomal markers from the Z-chromosome of chicken. It is further an object of the invention to use such markers to construct a Z-chromosome specific DNA map and to use such chromosomal markers to identify Z-chromosome homologs in related avian species, e.g., turkey.

In order to develop a dense genetic map for chicken, it is important to generate a large number of polymorphic markers per chromosome (Cheng et al, *Poultry Sci.*, 741:1855-1874 (1995)). One way of achieving this goal is to develop chromosome-specific libraries. Chromosome flow-sorting has been the method of choice for the generation of chromosome-specific libraries in humans (Fuscoe et al, *Cytogenet Cell Genet*, 43:79-86 (1986)) and in swine (Langford et al, *Anim. Genet*, 24: 261-267 (1993)). Development of flow-sorted chromosomes is technically demanding and frequently yield preparations which have some degree of contamination with other chromosomes (Hozier and Davis, *Anal. Biochem*, 200: 205-127 (1992)).

A more effective and direct way of generating chromosome-specific DNA libraries is by chromosome microisolation and microcloning of the chromosome of interest. Chromosome specific libraries generated by chromosome microisolation have been used in swine (Ambady et al, (unpublished data)), cattle (Ponce de León et al, *Proc. Natl. Acad. Sci., USA*, (in press) 1996)), and chicken (Li et al, *Proc. of the 10th Eur. Colloq. on Cytogenetics of Domestic Animals*, Utrecht Univ., The Neth., p. 11, August 18-21 (1992)) genetic mapping studies in order to develop maps for particular chromosomes.

Generation of polymorphic markers from chromosome-specific libraries for all of the 8 pairs of the chicken macrochromosomes will enable saturation of about 55-70% of the chicken genome. Chromosome-specific DNA can also be used as heterologous chromosome painting probes in closely and distantly related species for comparative genome analysis, study of chromosomal evolution, and for identifying gross chromosomal abnormalities.

This application, in particular, provides a chicken Z-chromosome-specific DNA library, Z-chromosomal markers and use thereof as probes to identify the Z-chromosome homolog in related species, such as turkey.

Brief Description of the Figures

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Figure 1 shows amplification of microsatellite markers by PCR and identification of polymorphisms.

Figure 2 shows a genetic map constructed using the identified microsatellite markers.

Figure 3 shows dinucleotide repeats present in the identified microsatellite markers.

Detailed Description of the Invention

Microisolation and microcloning:

Chicken metaphases were prepared from chicken fibroblast cultures following standard procedures, fixed briefly for 5 minutes each in 9:1, 5:1 and 3:1 methanol:acetic acid and dropped on clean coverslips. Chromosome microisolation and cloning was performed following the procedure described by Ponce de León et al (*Proc. Natl. Acad. Sci., USA* (in press) (1996)). Briefly, twelve copies of the chicken Z-chromosome were microisolated and transferred to clean siliconized coverslips. Proteinase-K digestion, phenol-chloroform extraction, *Sau*3AI (50U/ μ l, New England Biolabs) digestion and ligation to custom prepared *Sau*3AI adaptors were performed in a nanoliter drop. Ligation

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products were digested with BgII enzyme (Promega, 10 units/ μ l) to cleave off the adaptor dimers that form during the ligation process.

The ligation product was PCR amplified and 10 μ I of the amplified product was run on an agarose gel to determine the size of the amplified products. A 2 μ I volume of this original amplification was labeled by PCR, using biotin-16-dUTP (Boehringer Mannheim). The purity, specificity and origin of the DNA fragments was determined by FISH on chicken metaphases following the procedure described by Ponce de León et al (*Proc. Natl. Acad. Sci. USA* (in press) (1996)). The remainder of the PCR product was digested with *Sau3AI* and passed through a Microcon 30 (Amicon Inc.) spin column to cleave and remove the flanking adaptors respectively.

In order to produce a chicken Z-chromosome-specific phage library, the digested DNA was cloned in a lambda ZAP Express vector (Stratagene) and packaged using Gigapack II Gold packaging extract (Stratagene). The library was amplified by plate lysate method following the manufacturer's protocol and stored at -70°C in 7% DMSO and 0.3% chloroform. Average size of library inserts was determined by PCR amplification of 30 randomly picked clones using the T3 and T7 priming sites flanking the insert.

Fluorescent in situ hybridizations

The Z-chromosome-specific DNA fragments were fluorescently labeled by PCR with biotin-16-dUTP (3:1 ratio of dTTP:biotin-16-dUTP) and passed through a Sephadex G-50 column to remove unincorporated nucleotides. The protocol described by Ponce de León (*Proc. Natl. Acad. Sci., USA* (in press) (1996)) was followed. Briefly, 200 nanograms of labeled Z-chromosome specific DNA was mixed with 6 μ g of chicken competitor DNA (average size 200-400 bp) and 5.8 μ g of salmon sperm DNA (average size 200-400 bp), precipitated and resuspended in 12 μ l of hybridization buffer consisting of 50% deionized formamide, 1X SSC and 100% dextran sulphate to achieve a final DNA concentration of 1 μ g/ μ l. The hybridization mix was denatured at 75°C for 5

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minutes and reannealed at 37°C for 10 minutes and deposited on denatured (70% formamide, 2X SSC at 70°C for 2 minutes) chicken or turkey metaphases, mounted, sealed with rubber cement and incubated in a humidified chamber at 37°C for 18 to 20 hours. The slides were washed in 50% formamide/2X SSC at 42°C for 15 minutes and 0.1X SSC at 60°C for 15 minutes. Blocking was done using 2% blocking reagent (Boehringer Mannheim) and the signals were detected using avidin-FITC (5 μ g/ml, Vector labs) in 1% blocking solution. Slides were washed in 4X SSC/0.1% Tween-20 for 15 minutes at 42°C, stained for 10 minutes in propidium iodide (400 ng/ml in 2X SSC) and rinsed for 5 minutes in 2X SSC/0.01% Tween-20. Slides were mounted in p-phenylenediamine-11 (PPD-11) antifade and observed under a Zeiss Axioskop fluorescent microscope.

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Results

A chicken Z-chromosome specific DNA cocktail was developed by chromosome microisolation, Sau3AI digestion, adaptor ligation and PCR amplification. The amplified DNA fragments ranged in size from 400 bp to 1600 bp with the bulk of the DNA in the 500-1000 bp range. The origin, specificity and purity of the chromosomal DNA fragments was verified by FISH after PCR labeling of a small fraction of the DNA cocktail. The probes showed specific hybridization signal on a medium sized submetacentric chromosome identified as the Z-chromosome based on its morphology and G-banding pattern. After having confirmed the origin and purity of the preparation, the adaptors flanking the inserts were removed by Sau3AI digestion and column purification. Cloning was performed using equimolar ratios of the inserts to the vector ends (lambda ZAP Express, Stratagene). The original library consisted of a total of 8.48 X 10⁵ plaques representing about 14 chicken Z-chromosome equivalents. The final titer of the amplified library was 1.2 X 10¹² pfu/ml.

Thirty random plaques were selected and the inserts PCR-amplified using the T3/T7 priming sites flanking the inserts. The average insert size was about 1,000 bp (data not shown). This library was screened to identify microsatellite

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containing clones to increase the marker density of the chicken Z-chromosome genetic linkage map.

Heterologous painting of turkey metaphase chromosomes:

The labeled chicken Z-chromosome-specific DNA fragments were used to perform FISH analysis on turkey metaphase chromosomes following the procedure described previously. Washes at the same stringency showed strong hybridization signals on a medium-sized submetacentric chromosome in turkey metaphases (data not shown). This chromosome was identified as the Zchromosome homolog in the turkey. The obtained results indicate that the chicken and turkey Z-chromosome sequences are highly conserved. The redlegged partridge Z-chromosome has also been shown to be homologous to the chicken Z-chromosome (Dias el al, Proc. of the XXIV Int. Cont. on Anim. Genet., Prague, Czech. p. 133 (July 23-24, 1994)). These results are similar to the FISH results obtained when the bovine X-chromosome painting probes were used on sheep and goat chromosomes (Ponce de León el al, Proc. Natl. Acad. Sci., USA (in press) (1996)) and with human X-chromosome probes on a wide range of mammalian species (Schertan el al, Nat. Genet., 6:342-347 (1994)) indicating the high degree of sex chromosome conservation among all the mammalian species studied. Solinas-Toldo et al (Genomics, 27: 489-496 (1995)) have previously shown that human chromosome-specific painting probes could identify chromosomal segments in bovine that are homologous to specific human chromosomes. It is expected based on our results that chicken chromosome painting probes can similarly be used in closely and distantly related avian species to identify gross chromosomal rearrangements such as translocations and duplications that have occurred during avian evolution. Since the chicken Zchromosome sequences are highly conserved in the turkey, the chicken Zchromosome-specific microsatellite markers should be particularly useful for genetic mapping in turkey.

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Conclusions

Genetic and physical mapping of human and animal genomes has been greatly facilitated by the use of chromosome specific DNA libraries. Mapping with libraries specific to a chromosome or chromosomal region increases marker saturation by reducing the gaps resulting from a purely random shotgun approach. This study was undertaken to construct a genetic and physical map of microsatellites on the chicken Z chromosome. This chromosome is the fifth largest in the chicken genome, comprising about 8% of the total. Notwithstanding its size, very few microsatellites have been assigned to it. DNA originating from the chicken Z chromosome was previously isolated and reported. This was used to construct a small insert library in Lambda ZAP Express, representing 14 chromosome equivalents. This library was screened for microsatellites with an (AC) 12 oligo, and positive clones were isolated. Confirmation of the presence of the microsatellite, as well as its approximate location along the cloned fragment was accomplished by PCR amplification. Clones with adequate flanking regions were sequenced, and primers for 19 microsatellites were constructed. These primers were used to genotype individuals from the East Lansing Poultry Reference Population and a linkage map was constructed. Fourteen markers were scorable and polymorphic in this population. The resulting map contains 12 markers in two linkage groups spanning 90 Cm and two unlinked markers. The physical location of each marker was established by fluorescent in situ hybridization (FISH). Preliminary results with four markers allowed the assignment of one linkage group to the long arm of the Z chromosome, and one to the short arm.

The following nucleic acid sequences are microsatellite markers identified by the above methods. As discussed supra, these markers are useful for genetic mapping and for study of the sex chromosome structure in avian species. Also, such markers should enable the identification of genes encoding desirable traits, e.g., genes involved in growth rates, and for identifying sex-linked genotypes.

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EXAMPLE

The specific <u>Gallus domesticus</u> microsatellite markers identified are set forth below. As noted, these DNA markers will be useful for genetic mapping of domestic chicken as well as related avian species and for studies pertaining to evolution of the sex chromosome in avian species.

SEQUENCE 1 (43. Seq)

- 1 gateaettte cetaatatte ttgtgtttet tgtttgttga cetgtaatge
- 1 agttctgagt tttggaaagg aactaattaa gaccagagga gagataattt
- 101 tettttatea aaaaacaaac aaacaaacaa aaaaacgaat tettaccact
- 10 151 ttacaaaaat tttccatttt gaaggccagt acagccatag cattcatcta
 - 201 ctttttgctt tggat

SEQUENCE 2 (71. Seq)

- 1 gatcaggtgg cetgtagtag acaacaacaa caatggggtg cectttgttg
- 51 cettagtete taactegeae eeacacaca ttteaagttg ettgtggeea
- 15 101 ttetteaggg acagttette acaatetatt cettteetga tgtagaagge
 - 151 gtcacctcct cccctcctgc ctcgtttgtc ccttctaaac tgcaggtatt
 - 201 agtattgata getaaggtea agteatggga accateteae eaggttteag
 - 251 tgttggcaac tatgttatgc tttcttagga gcatggtggt tccaactctt
 - 301 ccctgcttat ttcccaagct gtgtgtgatg gtaggatagc attcaagtgg
- 20 351 gaggageeta teggettttt ggaggtaete etaaateeet gatatteeee
 - 401 tgattecegt aettetteet tgeeaaggge eegeeaatge atagtteaat
 - 451 tteteatgea gaegetaagg aaaggtggae ee

SEQUENCE 3 (80 Seq.)

- 1 gategtatgt attttttac ataggataga aaatggccaa taggaaataa51 gacagtacag etactaagaa agaaacacaa ttacacacac acacacacac
- 101 acacacaca acacatttga aaaacgcgct gcacagcagt gtgggtattt
- 5 151 tttcacaaga gagacacact ctacagtaca cagccagctc tactttgtcg
 - 201 cacagtetea gtgtgtgttt gecaacagga egeggtteae agggagatat
 - 251 tgtcctcttg tgtgtgtgga gacacagaga cagag

SEQUENCE 4 (81. Seq)

- 1 gateeeetgg aggaagggea atggeaacee acteeagtat tettgeetga
- 10 51 agaataccat ggtcagtttt gcctcctggg ctatagtcca tggggttgca
 - 101 aagagteagg catgactgag egactetete tetetetete tetetetete
 - 151 acacacaca acacacaca acacacggeg tetetetete tetetataca
 - 201 tataggetgt gtgteteget atteteacat gagggaaaet catatetage
 - 251 acgtggcaca aatattgttt gtggctctca caaaagacat gtgggcgcac
- 15 301 aaaggteece eeeeggtgga taeanegeet tggtttttta taaceeaage
 - 351 ctgtg

SEQUENCE 5 (131 Seq)

- 1 gatcacatat gtaaactagg gaattgcata ataagattaa atgtaggtgt
- 51 agaacgtggc atgaaggaag gtagaattag gtggtaccta tctcttctga
- 20 101 aacaaactga gaatcctact accaatcaac atattetaca taccacacac
 - 151 acattttttc tcgagtaaaa tataaactaa tgagaaactt ccctag

SEQUENCE 6 (147. Seq)

	1	gateceaage aacacatagn cagacaatca cacacacaca cacacacaca
	51	cacacacaca cacacacaca cacatectet ecceacaata cateeegaga
	101	ggggggagag acaetetete teceteteta taggggagae ceggagage
5	151	ggetetgttg tetetetaca eeggacatae agtggageae ateteacaet
	201	tgtgtctttg tetetetaea eeggacatae agtggageae ateteaeaet
	251	tgtgteteta teteteeetg teeetgttga teeatetete tteacacate
	301	tetecagate ttagegetag agteteetgt ettetetetg egeaatttgt
	351	gtgatagaga cacctgatat gttgtgtggg ggagacatct gtgtgtctct
10	401	gtgtcatccc agaggatttt tctctcccac acttagagge cttctcaaga
	451	gatgggaggt tttaatgggg tgtg

SEQUENCE 7 (166. Seq)

gateattett etgttteeea ttetaatggg aatteteeae acacacac
 acacacacac acacacacat ettetteeee ttacatggaa aaaaateete
 cacaccectg gacactgatt acteteeete tteeeagaga gagate

SEQUENCE 8 (196. Seq)

1 gatecectag agaagggaat ggetacteae teeagtatte ttgeetggag
51 aatteegtgg teagaggage etggaagget ataateeata gagtegeaag
101 agteagacag gaetgagtga etaacacaca eatgeacaca cacacacaca
20 151 cacacacaca ettgetetag ggagaggeat agagatgtaa teteteetaa
201 aatgggggtg gegatggeee etgeggeeaa gtaategeea eacatgegta
251 tteeeettaa gattgggtta ggeeteeett atgaggagag accagggaga
301 gaatgggete tetetetete teaeteeeca accgagtaag tggtaaaaaa
351 ggtttteetg gattacaatt ttggtgttae agaattggaa aaaaatattt
25 401 ttggggetee eeeeteagtt ta

SEQUENCE 9 (199. Seq)

- 1 ctagcaaaaa caccccaca agttatgaaa acaacggctt aatatagtaa
- 51 tgtgtgtgt tgtgtgtgt tgttgcacac cacagttttc tctgatactc
- 101 aaacetetet etttetetae aggggeeece eataacacag eggetgagat
- 5 151 gtgtgacggg aaggcgtggc cttttacaca tttgtggtat ggtctgccaa
 - 201 ggececetat tgecececae aactaeggag atacaetagg ggegaeeege
 - 251 aggegegega ecceeaggtg gggeeeegag

SEQUENCE 10 (204. Seq)

- 1 etttaggagg ttetetegag taagettttt ggatttettt ggtteecaag
- 10 51 catcacatgg tacaggcagt cacacacaca cacatacaca cacacacaca
 - 101 cacacacaca cacteetete eccacaatac atacegagag gggggagaga
 - 151 cactetetet ecetetetat agggggagee ceaeagaget ggetetgttg
 - 201 tetetecea eeggacatae agtggageae ateteacaet tetgteteta
 - 251 tetetecetg eccetgtgae atceatetet etteacacaa teteacecag
- 15 301 gatettageg etagagaeee eetgteette tteteetggg gaaatttttt
 - 351 gtggataaga gacacccgat atattggtgt gggggagaac atcttgtgag
 - 401 gtetetgttg tgeeateeca acaggaattt ttateteece cacaattaga
 - 451 ggcccctcct caagagtgtg tgagggtt

SEQUENCE 11 (235. Seq)

- 20 1 gatcacagat gtatgtattt ttttacatag gatagaaaat ggacaatagg
 - 51 aaataagaca gtacagctac taagaaagaa cccacattta cacacacaca
 - 101 cacacacaca cacacacaca agtgtttaat ccgctgcaca gcattgtgga
 - 151 catttttaca caagagagac acactctaca gtttgcgccc agctctag

SEQUENCE 12 (249. Seq.)

- 1 gateattett etgttteeea ttetaatgga atteteeaea eacacacae
- 51 cacacacaca cacacactet tettteteet gacatggaaa aateteecee
- 101 acacceggg acactgattt etetecetet eeceaacaet gtgagcaaga
- 5 151 ggagtttatt ttgtgtgtgt cactetteca gggagagaga gate

SEQUENCE 13 (258. Seq)

- 1 ctaggcateg gttgggaggt ggtgagtaat tacttgtetg acattagtee
- 51 tgtaacattg ggtgtgtgtg tgtgtgtgtg tgtgtattcc ccttgggaat
- 101 tggttttete aaccacaagt tettetttt tttttttete ecceetttte
- 10 151 ttctgaaaat aagtacttgg ggggtttccg cccccccgg taaataaaat

SEQUENCE 14 (290. Seq)

- 1 ctagtggete ecaageaaca catageeaga caacacaca acacacaca
- 51 acacacaca acacacaca acacacacte etetececae aatacateee
- 101 gagaggggg agagacactc tetetecete tetatagegg gageceeaca
- 15 151 gagetggete tgetgtetet etaeaeegga eataeagtgg ageaeatete
 - 201 acattegtgt etetatetet eeetgeeeet ggtgacatae atetetette
 - 251 acacatetea ceaggtetga gegetagagt etcetgtett etetetgege
 - 301 aatatttgtg atagagacat ctgatatatt gtgtgtggga gacatettgt
 - 351 gagtetetgt gtgeatecea gaggattttt ateteeceae aetag

SEQUENCE 15 (309. Seq)

- 1 gatccatgaa aactttccga gttgtattgt ctaggtgaaa acacacacaa
- 51 acacacaca acacacaca acacaacagg gagatgagtc ttgcaagaga
- 101 ataggggaga gttatgtcac caagtctggt gaggtatata gcgtataggg
- 5 151 agccaacatg teagacatet gatgtgetaa gattaacatt ttattttatt
 - 201 taatgtgtga gatctcatat ageggetett ettatatatg aegtetegea
 - 251 atgtetettt atgtgtgtta ttetetgage eeetgggaga tatetgteat
 - 301 cagagagaag agacatacac atacaggggt tatatatttt ctccctgtgt
 - 351 gtggagatgg agggtatttt ggacaagete aacaeteatt ggeteecaga
- 10 401 gagagaaaag gagcaactgt tgcacccggg gctctgtagc tgggatc

SEQUENCE 16 (341. Seq)

- 1 caattgggta catctacctg gtaccccacc cgggtggaaa atcgcatggg
- 51 ceegeggegg ttetaggaag taetetegag aagettttgg gttetttggg
- 101 teccaageag cacatggaca ggeaateaca cacacacaca cacacacaca
- 15 151 cacacacaca cacacacaca etceteteee cacaatacat eeegagaggg
 - 201 gggagagtea etetetetee etetetatag ggggegeece taagagetgg
 - 251 ctetattate tatetacace geacatacaa tagageacaa eteacactag

SEQUENCE 17 (398. Seq)

- 1 gatcaaagca tggaggtcat gccaggcact gaacaaaatg gtagagagtg
- 20 51 attetatgae tgaetaagae eteatgeaae aacaagtgaa gagteacaae
 - 101 tgcaaacaga agtacaactt agcaaatcct attttcagga aacactaaac
 - 151 cgtaatactt gcacgatttt ttctttaata cagtaataat tcttttagaa
 - 201 tttggatata tettttaaga taeatatttg tetaaataee aaggeaggat
 - 251 atgagcataa aatagctaag gttagctatg gtgttatatt taagaagacc
- 25 301 acagagcaat aggagcatac ttttcttggg gtagaagggg cccttaaagg
 - 351 teacetag

SEQUENCE 18 (420. Seq)

- 1 ctagecaeat cetataaete eaeteeaeet ttaateetga tttetgtgte
- 51 tettetetaa eetetatgge etttetetaa agtteeccaa tateaacaat
- 101 ccttttcccc actgggacct ccagtttatt gattctacca tgtcactatc
- 5 151 catggtcaac cacttgtggt attataggat gtcgcgtgtg tgtgtgtgtg
 - 201 tgtgtgcatg tgtgtgtgct tgggtgtcag agagttccaa tctgggggac
 - 251 ctatggtttg taaacaacag gtetettgee aaggaagat

SEQUENCE 19 (435. Seq)

- 1 ctagegeteg tgeecetgea gttegacaet eagtggetee teeacaeaea
- 10 51 cacacacaca cacatcaata tatatataga tagatagata gatagaggag
 - 101 caatataagt ggetteteta tttecageat gttttgaaga geataaacte
 - 151 aacagagtat atataaatet gatgtgaccc atgtcatetg ctacagcatg
 - 201 agagggggta gtgatc

CLAIMS:

- 1. A Z-chromosomal marker DNA selected from the group consisting of Sequence I (43. Seq), Sequence 2 (71. Seq), Sequence 3 (80. Seq), Sequence 4 (81. Seq), Sequence 5 (131. Seq), Sequence 6 (147. Seq), Sequence 7 (166. Seq), Sequence 8 (196. Seq), Sequence 9 (199. Seq), Sequence 10 (204. Seq), Sequence 11 (235. Seq), Sequence 12 (249. Seq), Sequence 13 (258. Seq), Sequence 14 (290. Seq), Sequence 15 (309. Seq), Sequence 16 (341. Seq), Sequence 17 (398. Seq), Sequence 18 (420. Seq), and Sequence 19 (435. Seq).
- 2. A Z-chromosomal DNA library that contains at least one DNA sequence according to Claim 1.
 - 3. A method of using at least one Z-chromosomal DNA according to Claim 1 for genetic mapping.
 - 4. The method of Claim 3, wherein the genetic mapping is effected to construct a Z-chromosome specific DNA map.
- 5. The method of Claim 3, wherein the Z-chromosome DNA map is that of an avian species selected from the group consisting of chicken, turkey, partridge, duck, guinea hen, and goose.
 - 6. The method of Claim 4, which is used to identify gross chromosomal rearrangements.
- 7. The method of Claim 6, wherein said chromosomal rearrangement comprises a translocation, deletion or duplication.

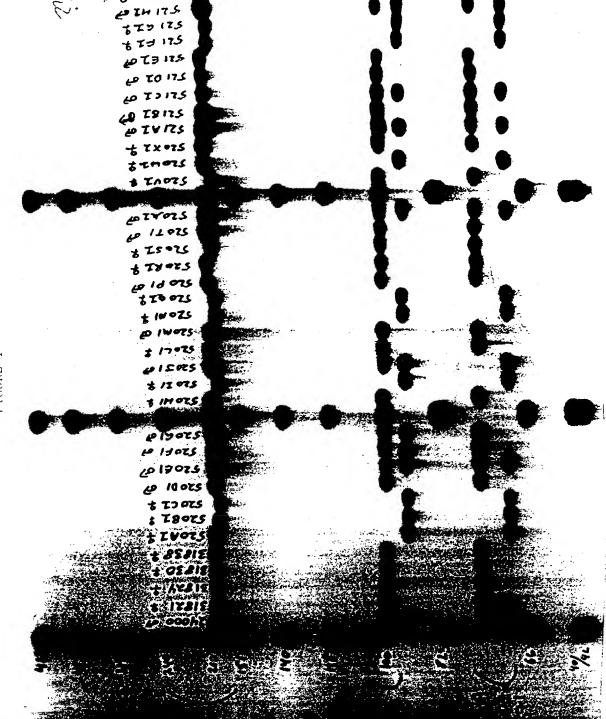


FIGURE 1

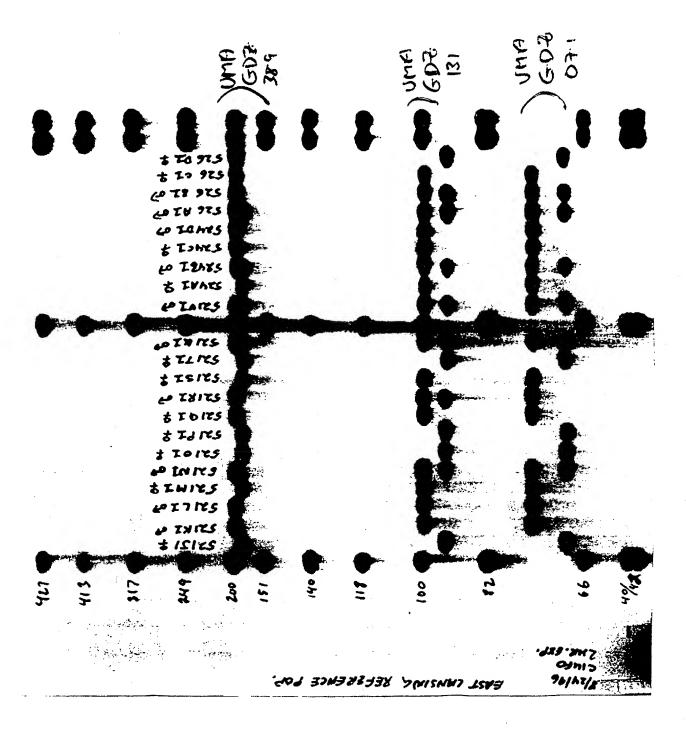
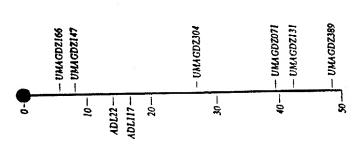
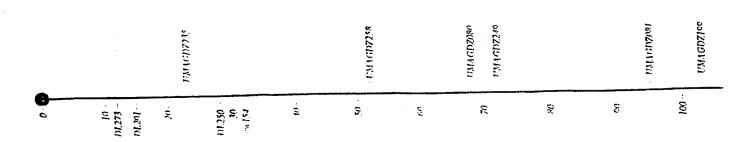


FIGURE 1 (Cont)

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FIGURE 2





Chicken Z Chromosome Microsatellites Microsatellite composition

s. Ciufo

Clone	Repeat
UMGDZ043	(AAC) ₇
UMGDZ071	(CA) ₅
UMGDZ080	(AC) ₁₆
UMGDZ081	(CT) ₁₃ (AC) ₁₃ (CT) ₇
UMGDZ131	(CA) ₄
UMGDZ147	(CA) ₂₂
UMGDZ166	(AC) ₁₅
UMGDZ196	(AC) ₁₉
UMGDZ199	(GT) ₁₂
UMGDZ204	(AC) ₂₁
UMGDZ235	(AC) ₁₅
UMGDZ249	$(AC)_{16}(TTC)_4$
UMGDZ258	(TG) ₁₂
UMGDZ290	(AC) ₂₃
UMGDZ304	(AC) ₂₀
UMGDZ341	(AC) ₂₂
UMGDZ398	(CAA) ₃
UMGDZ420	(GT) ₂₀
UMGDZ435	(CA) ₁₁

FIGURE 3

cional Application No

PCT/US 98/08896 CLASSIFICATION OF SUBJECT MATTER IPC 6 C1201/68 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC 6 C12Q Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Relevant to claim No. Category Citation of document, with indication, where appropriate, of the relevant passages LEVIN I ET AL: "Genetic map of the 1 - 7Α chicken Z chromosome using random amplified polymorphic DNA (RAPD) markers." GENOMICS, (1993 APR) 16 (1) 224-30, XP002067078 cited in the application see the whole document WO 94 07907 A (ZOOGEN INC) 14 April 1994 1-7 Α see the whole document WO 96 39505 A (ISIS INNOVATION ; GRIFFITHS 1 - 7Α RICHARD (GB); TIWARI BELA (GB)) 12 December 1996 see the whole document Further documents are listed in the continuation of box C. Patent family members are listed in annex. Special categories of cited documents: "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another "Y" document of particular relevance; the claimed invention citation or other special reason (as specified) cannot be considered to involve an inventive step when the "O" document referring to an oral disclosure, use, exhibition or document is combined with one or more other such doc ments, such combination being obvious to a person skilled in the art. document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of the actual completion of theinternational search Date of mailing of the international search report 4 June 1998 18/06/1998 Name and mailing address of the ISA Authorized officer

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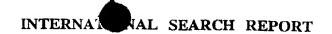
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Molina Galan, E



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